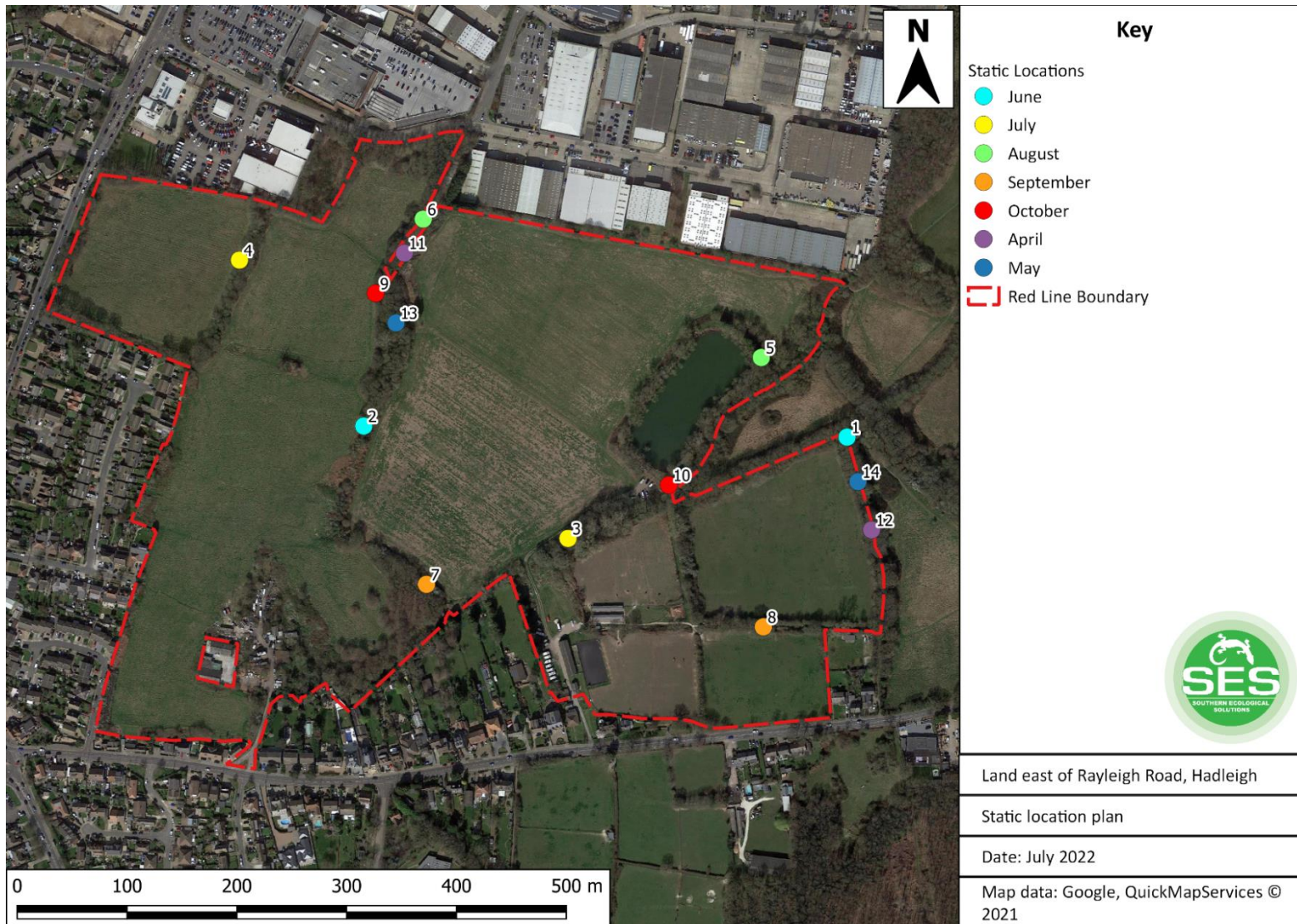


Appendix 6h – Static Detector Locations



Appendix 6i – Activity Transect Survey Results and Heat Maps

June 2020

Table A7.2: Summary of June 2020 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
24/06/2020	June dusk transect	Start/Sunset: 21:20 Finish: 23:20	20°C 0% Cloud 3 Beaufort	Anti-clockwise
Time	Comments			
21:20-21:25				
21:25-21:30				
21:30-21:35	4x Soprano pipistrelle			
21:35-21:40				
21:40-21:45				
21:45 – 21:50				
21:50 – 21:55				
21:55 – 22:00	1x Soprano pipistrelle			
22:00 – 22:05				
22:05 – 22:10				
22:10 – 22:15	1x Common pipistrelle, 1x Soprano pipistrelle			
22:15 – 22:20	4x Soprano pipistrelle			
22:20 – 22:25	2x Common pipistrelle, 1x Soprano pipistrelle, 1x Daubenton's bat			
22:25 – 22:30	2x Common pipistrelle			
22:30 – 22:35	2x Common pipistrelle			
22:35 – 22:40	3x Common pipistrelle, 1x <i>Myotis sp.</i> Daubenton's bat			
22:40 – 22:45	3x Common pipistrelle			
22:45 – 22:50				
22:50 – 22:55				
22:55 – 23:00				
23:00 – 23:05	1x Noctule			
22:05 – 23:10	2x Common pipistrelle			
23:10 – 23:15	2x Common pipistrelle			
23:15 – 23:20	4x Common pipistrelle, 2x Soprano pipistrelle, 1x Noctule, 1x Daubenton's bat			

July 2020

Table A7.3: Summary of July 2020 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
21/07/2020	July dusk transect	Start/Sunset: 21:02 Finish: 23:07	19°C 20% Cloud 0 Beaufort	Clockwise
Time	Comments			
21:02 – 21:07				
21:07 – 21:12				
21:12 – 21:17				
21:17 – 21:22				
21:22 – 21:27				
21:27 – 21:32				
21:32 – 21:37	1x Soprano pipistrelle			
21:37 – 21:42				
21:42 – 21:47	1x Common pipistrelle, 1x Soprano pipistrelle			
21:47 – 21:52	2x Common pipistrelle, 2x Soprano pipistrelle			
21:52 – 21:57	1x Soprano pipistrelle			
21:57 – 22:02				
22:02 – 22:07	2x Daubenton's bat			
22:07 – 22:12	2x Common pipistrelle, 1x Noctule			
22:12 – 22:17	1x Common pipistrelle, 4x Soprano pipistrelle			
22:17 – 22:22	1x Soprano pipistrelle			
22:22 – 22:27	2x Soprano pipistrelle, 1x Daubenton's bat			
22:27 – 22:32	1x Common pipistrelle, 4x Soprano pipistrelle			
22:32 – 22:37	2x Soprano pipistrelle, 1x <i>Nyctalus sp.</i>			
22:37 – 22:42	2x Common pipistrelle, 4x Soprano pipistrelle			
22:42 – 22:47	5x Common pipistrelle, 4x Soprano pipistrelle			
22:47 – 22:52	3x Common pipistrelle, 2x Soprano pipistrelle, 2x Brown long-eared bat			
22:52 – 22:57	1x Soprano pipistrelle			
22:57 – 23:02	1x Soprano pipistrelle			
23:02 – 23:07	2x Soprano pipistrelle			

August 2020

Table A7.4: Summary of August 2020 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
06/08/2020	August dusk transect	Start/Sunset: 19:56 Finish: 22:04	19°C 90% Cloud 1 Beaufort	Anti-Clockwise
Time	Comments			
19:56 – 20:01				
20:01 – 20:05				
20:05 – 20:10				
20:10 – 20:21	1x Soprano pipistrelle			
20:21 – 20:26	2x Common pipistrelle			
20:26 – 20:29				
20:29 – 20:34	2x Soprano pipistrelle			
20:36 – 20:41	1x Common pipistrelle, 2x Soprano pipistrelle			
20:41 – 20:46	2x Common pipistrelle, 1x Daubenton's bat			
20:46 – 20:52	1x Common pipistrelle			
20:52 – 20:57	1x Soprano pipistrelle			
20:57 – 21:04	1x Common pipistrelle, 2x Daubenton's bat			
21:04 – 21:09	1x Noctule			
21:09 – 21:16	6x Common pipistrelle			
21:16 – 21:21	1x Common pipistrelle, 2x Brown long-eared bat			
21:21 – 21:46	1x Noctule, 1x Daubenton's bat			
21:46 – 21:51				
21:51 – 22:04				

September 2020

Table A7.5: Summary of September 2020 dusk bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
22/09/2020	September dusk transect	Start/Sunset: 18:55 Finish: 20:55	20°C 30% Cloud 2 Beaufort	Clockwise
Time	Comments			
18:55 – 19:00				
19:00 – 19:05				
19:05 – 19:10				
19:10 – 19:15	1x Common pipistrelle			
19:15 – 19:20	1x Soprano pipistrelle			
19:20 – 19:25	1x Soprano pipistrelle			
19:25 – 19:30	2x Common pipistrelle			
19:30 – 19:35	1x Soprano pipistrelle			
19:35 – 19:40				
19:40 – 19:45				
19:45 – 19:50	1x Common pipistrelle			
19:50 – 19:55				
19:55 – 20:00				
20:00 – 20:05	2x Common pipistrelle			
20:05 – 20:10				
20:10 – 20:15				
20:15 – 20:20				
20:20 – 20:25	2x Common pipistrelle			
20:25 – 20:30				
20:30 – 20:35				
20:35 – 20:40	2x Common pipistrelle			
20:40 – 20:45	1x Common pipistrelle			
20:45 – 20:50	2x Common pipistrelle			
20:50 – 20:55	1x Common pipistrelle			

Table A7.6: Summary of September 2020 dawn bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
23/09/2020	September dawn transect	Start/Sunset: 04:46 Finish: 06:46	16°C 100% Cloud 0 Beaufort	Anti-Clockwise
Time	Comments			
04:46 – 04:51	2x Common pipistrelle			
04:51 – 04:56	1x Common pipistrelle			
04:56 – 05:01				
05:01 – 05:06	1x Common pipistrelle			
05:06 – 05:11				
05:11 – 05:16				
05:16 – 05:21	1x Common pipistrelle			
05:21 – 05:26	1x Common pipistrelle			
05:26 – 05:31	1x Common pipistrelle, 1x Soprano pipistrelle			
05:31 – 05:36	2x Common pipistrelle			
05:36 – 05:41				
05:41 – 05:46	1x Common pipistrelle			
05:46 – 05:51	1x Common pipistrelle			
05:51 – 05:56	1x Common pipistrelle, 1x Soprano pipistrelle			
05:56 – 06:01				
06:01 – 06:06				
06:06 – 06:11				
06:11 – 06:16				
06:16 – 06:21				
06:21 – 06:26				
06:26 – 06:31				
06:31 – 06:36				
06:36 – 06:41				
06:41 – 06:46				

October 2020

Table A7.7: Summary of October 2020 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
22/10/2020	October dusk transect	Start/Sunset: 17:52 Finish: 19:52	14°C 75% Cloud 1 Beaufort	Clockwise
Time	Comments			
17:52 – 18:05				
18:05 – 18:10				
18:10 – 18:24				
18:24 – 18:29				
18:29 – 18:36				
18:36 – 18:45	1x Common pipistrelle			
18:45 – 18:50	1x Common pipistrelle			
18:50 – 18:55				
18:55 – 19:00				
19:00 – 19:05				
19:05 – 19:13				
19:13 – 19:29	1x Common pipistrelle, 1x Soprano pipistrelle			
19:29 – 19:34				
19:34 – 19:40				
19:40 – 19:47				
19:47 – 19:52				

April 2021

Table A7.8: Summary of April 2021 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
27/04/2021	April dusk transect	Start/Sunset: 21:15 Finish: 22:30	10°C 70% Cloud 2 Beaufort	Clockwise
Time	Comments			
20:15-20:20				
20:20-20:25				
20:30-20:35				
20:35-20:40	1x Common pipistrelle			
20:40-20:45				
20:45 – 20:50	1x Common pipistrelle			
20:50 – 20:55	2x Common pipistrelle			
20:55 – 21:00				
21:00 – 21:05				
21:05 – 21:10	1x Daubentons bat			
21:10 – 21:15				
21:15 – 21:20	1x Daubentons bat			
21:20 – 21:25				
21:25 – 21:30	2x Daubentons bat			
21:30 – 21:35	1x Common pipistrelle			
21:35 – 21:40	4x Common pipistrelle, 1x Soprano pipistrelle, 1x <i>Myotis sp.</i> 1x Daubenton's bat			
21:40 – 21:45	1x Common pipistrelle			
21:45 – 21:50				
21:50 – 21:55				
21:55 – 22:00				
22:00 – 22:05	2x Soprano pipistrelle, 4x <i>Myotis sp</i>			
22:05 – 22:10	4x <i>Myotis sp</i>			
22:10 – 22:15				
22:15 – 22:20	2x Soprano pipistrelle			
22:20 – 22:25	2x Soprano pipistrelle			
22:25 – 22:30				

May 2021

Table A7.9: Summary of May 2021 bat transect results.

Date	Survey type	Survey timings	Weather	Route Direction
20/05/2021	May dusk transect	Start/Sunset: 20:51 Finish: 22:51	12°C 90% Cloud 4 Beaufort	Clockwise
Time	Comments			
20:50-20:55				
20:55-21:00				
21:00-21:05				
21:05-21:10				
21:10-21:15				
21:15 – 21:20				
21:20 – 21:25	1x Soprano pipistrelle			
21:25 – 21:30				
21:30 – 21:35				
21:35 – 21:40	1x Soprano pipistrelle			
21:40 – 21:45				
21:45 – 21:50				
21:50 – 21:55	1x Soprano pipistrelle, 1x Barbastelle			
21:55 – 22:00				
22:00 – 22:05				
22:05 – 22:10	6x Soprano pipistrelle, 6x Common pipistrelle			
22:10 – 22:15	6x Common pipistrelle, 7x Soprano pipistrelle, 1x Daubentons bat			
22:15 – 22:20	1x Soprano pipistrelle, 4x Daubentons bat			
22:20 – 22:25	11x Soprano pipistrelle, 1x Daubentons bat			
22:25 – 22:30	2x Common pipistrelle			
22:30 – 22:35				
22:35 – 22:40				
22:40 – 22:45	1x Common pipistrelle			
22:45 – 22:50	1x Soprano pipistrelle			
22:50 – 22:55				

Bat Activity Hotspots



Key

 Red Line Boundary



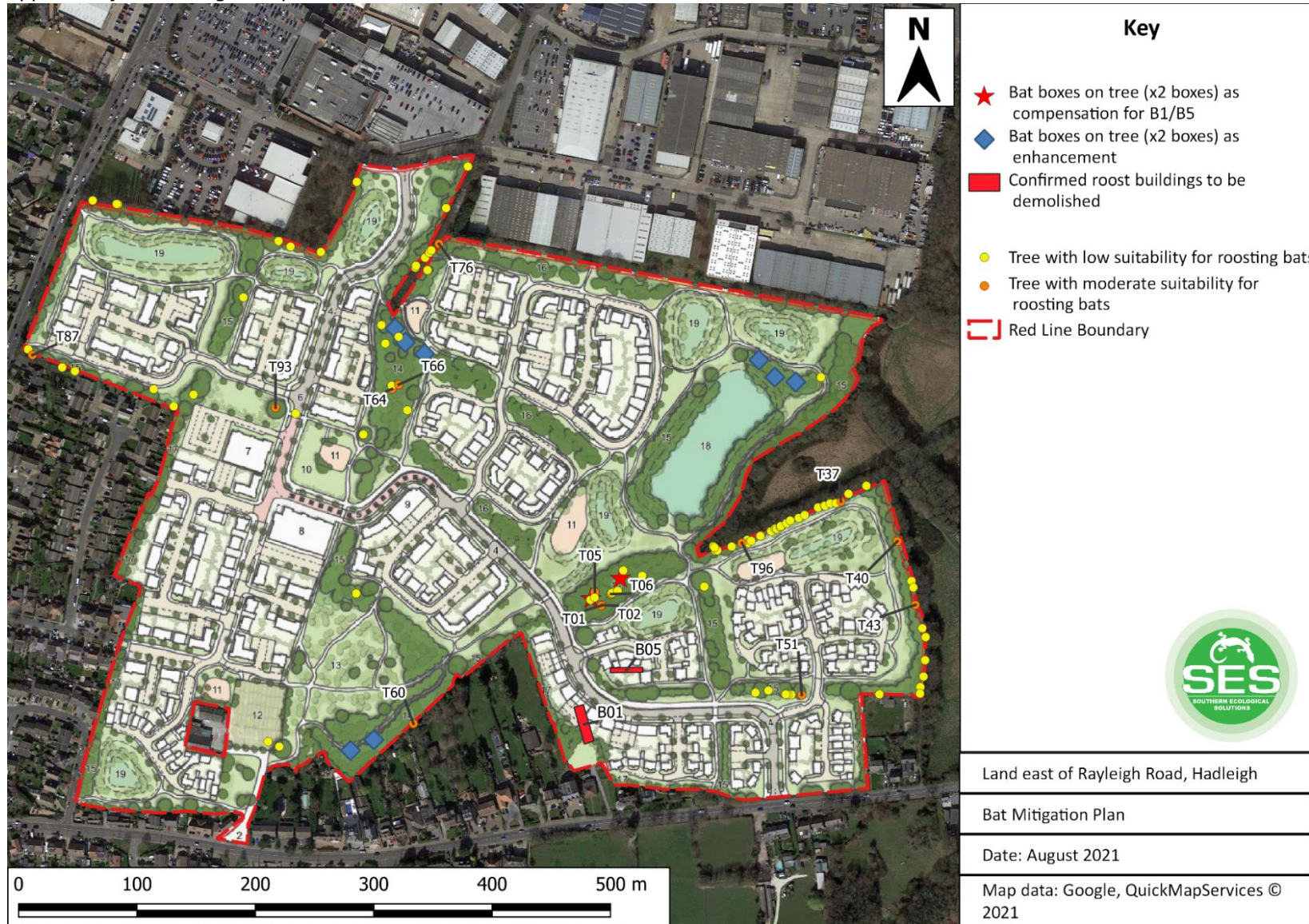
Land east of Rayleigh Road, Hadleigh

Bat Activity Hotspots

Date: July 2022

Map data: Google, QuickMapServices © 2021

Appendix 6j – Bat mitigation plan



Appendix 7: Wintering Bird Survey Results

Appendix 7a – Wintering Bird Survey Results and Weather Conditions

Table A8.1: Status of wintering birds within the site.

Species	BoCC	S. 41	V1	V2	Comments
Fieldfare <i>Turdus pilaris</i>	Red		1	10	Foraging in the paddocks and surrounding hedgerows.
Herring gull <i>Larus argentatus</i>	Red	✓	1	14	Foraging and shelter within the fields.
House Sparrow <i>Passer domesticus</i>	Red	✓	1	4	Low numbers within areas of scrub adjacent to residential development.
Starling <i>Sturnus vulgaris</i>	Red	✓	2	1	
Black-headed gull <i>Chroicocephalus ridibundus</i>	Amber		1	5	Flying over site and present on the lake
Dunnock <i>Prunella modularis</i>	Amber	✓	5	6	Associated with the hedgerows and scrub across the site.
Lesser black-backed gull <i>Larus fuscus</i>	Amber		0	2	
Mallard <i>Anas platyrhynchos</i>	Amber		3	3	Foraging and sheltering on the lake.
Wren <i>Troglodytes troglodytes</i>	Amber		0	1	
Redwing <i>Turdus iliacus</i>	Amber		1	25	Foraging in the paddocks and surrounding hedgerows.
Song thrush <i>Turdus philomelos</i>	Amber	✓	2	1	
Woodpigeon <i>Columba palumbus</i>	Amber		1	20	
Blackbird <i>Turdus merula</i>	Green		10	4	
Blue tit <i>Cyanistes caeruleus</i>	Green		3	8	
Buzzard <i>Buteo buteo</i>	Green		1	1	Hunting over site
Carrion crow <i>Corvus corone</i>	Green		7	2	
Chaffinch <i>Fringilla coelebs</i>	Green		1	2	
Chiffchaff <i>Phylloscopus collybita</i>	Green		0	1	
Cormorant <i>Phalacrocorax carbo</i>	Green		0	2	Hunting in the lake.
Coal tit <i>Periparus ater</i>	Green		0	1	
Coot <i>Fulica atra</i>	Green		2	1	Foraging and sheltering on the lake.
Great tit <i>Parus major</i>	Green		3	4	
Great spotted woodpecker <i>Dendrocopus major</i>	Green		1	0	
Grey heron <i>Ardea cinerea</i>	Green		0	1	Hunting in the lake.
Jay <i>Garrulus glandarius</i>	Green		2	2	Foraging on site.
Magpie <i>Pica pica</i>	Green		3	3	
Moorhen <i>Gallinula chloropus</i>	Green		1	0	Foraging and sheltering on the lake.
Pied wagtail <i>Motacilla alba</i>	Green		0	1	

Species	BoCC	S. 41	V1	V2	Comments
Robin <i>Erithacus rubecula</i>	Green		5	6	
Pheasant <i>Phasianus colchicus</i>	Introduced		0	1	

Red rows are BOCC red-list, Amber rows are BoCC amber-list, Green rows are BoCC green-list.
BoCC: Birds of Conservation Concern as defined and listed in Eaton *et al.*, (2021)

Table A8.2: Summary of wintering bird survey visit dates and weather conditions.

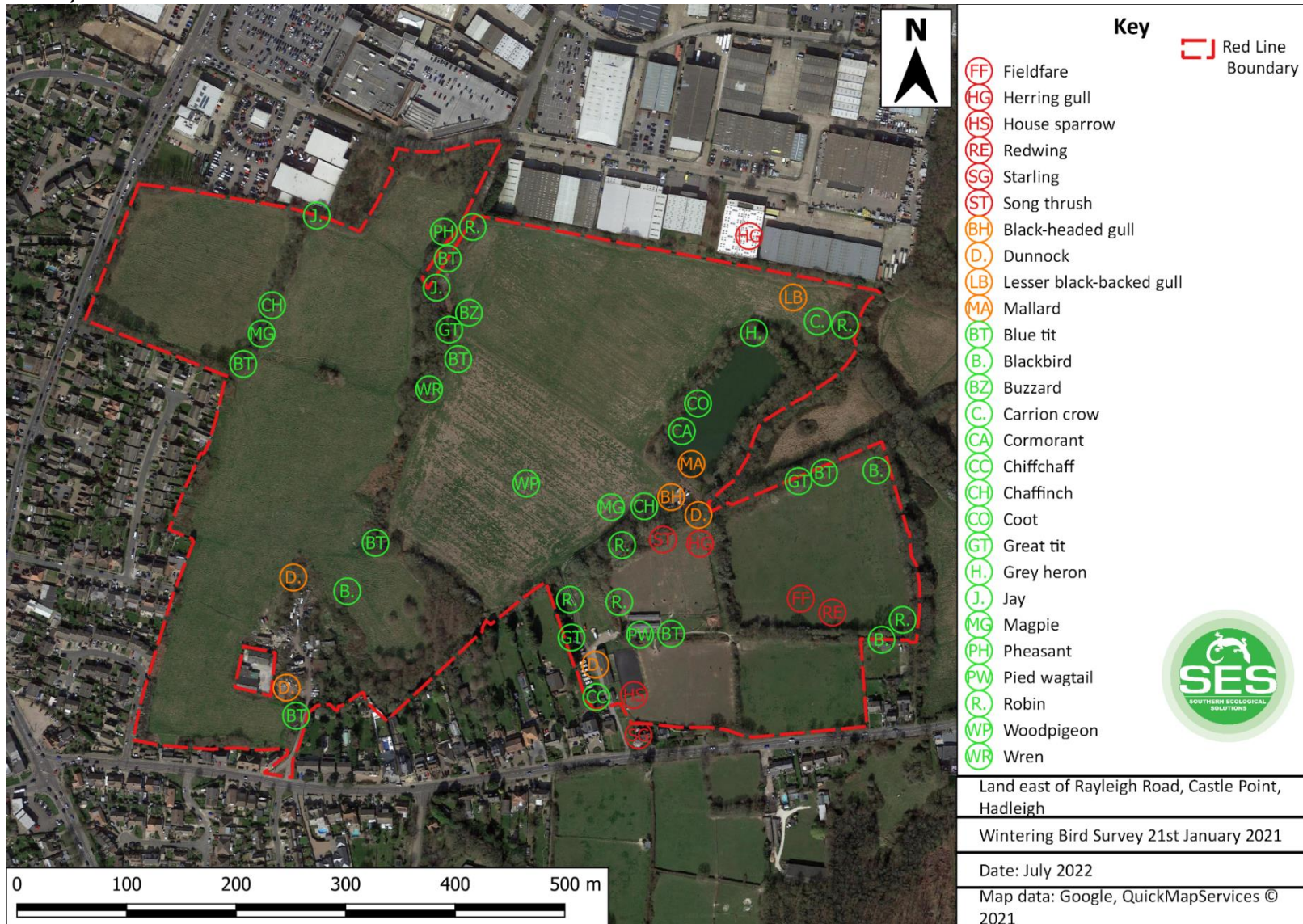
Visit	Date	Survey Conditions
1	23/12/2020	Fair: 11°C (average), little precipitation, 2 wind, cloud 8/8, good visibility.
2	21/01/2021	Fair: 6°C (average), no precipitation, 4 wind, cloud 1/8, good visibility.

Appendix 7c – Wintering Bird Survey Maps

December 2020



January 2021



Appendix 8: Breeding Bird Survey Results

Appendix 8a – Breeding Bird Survey Results and Weather Conditions

Table A9.1: Status of breeding birds within the site.

Species	BoCC	S. 41	V1	V2	V3	V4	Status
Herring gull <i>Larus argentatus</i>	Red	✓	1	0	2	1	Foraging on the site
House Sparrow <i>Passer domesticus</i>	Red	✓	5	16	10	14	Small colony adjacent to site and foraging in boundary hedgerows and scrub
Common linnet <i>Linaria cannabina</i>	Red	✓	1	0	6	0	Foraging on the site
Starling <i>Sturnus vulgaris</i>	Red	✓	2	0	4	7	Pairs in housing adjacent to site; foraging within site
Willow tit <i>Poecile montanus</i>	Red	✓	0	0	0	3	Foraging on the site
Woodcock <i>Scolopax rusticola</i>	Red		0	1	0	0	Foraging on the site
House martin <i>Delichon urbicum</i>	Red		0	1	0	0	Foraging over the site.
Greenfinch <i>Chloris chloris</i>	Red		0	0	2	1	Possible breeding - 1 territory
Song thrush <i>Turdus philomelos</i>	Amber	✓	1	2	1	0	Probable breeding – 1 territory
Black-headed gull <i>Chroicocephalus ridibundus</i>	Amber		0	0	1	0	Flying over site
Dunnock <i>Prunella modularis</i>	Amber	✓	7	2	4	1	Probable breeding – 2 territories
Green sandpiper <i>Tringa ochropus</i>	Amber		1	2	0	0	Foraging on site.
Kestrel <i>Falco tinnunculus</i>	Amber	✓	0	1	0	2	Hunting over site
Lesser black-backed gull <i>Larus fuscus</i>	Amber		0	0	0	1	Flying over the site
Mallard <i>Anas platyrhynchos</i>	Amber		3	0	4	0	Probable breeding – 2 territories
Stock dove <i>Columba oenas</i>	Amber		0	2	0	0	Flying over the site
Moorhen <i>Gallinula chloropus</i>	Amber		2	0	0	0	Foraging on site.
Woodpigeon <i>Columba palumbus</i>	Amber		1	1	0	0	Foraging on the site
Blackbird <i>Turdus merula</i>	Green		8	3	2	4	Probable breeding – 2 territories
Blackcap <i>Sylvia atricapilla</i>	Green		2	1	5	4	Probable breeding - 1 territory
Blue tit <i>Cyanistes caeruleus</i>	Green		15	2	14	8	Probable breeding - 3 territories
Carrion crow <i>Corvus corone</i>	Green		3	0	2	0	Foraging on site
Chaffinch <i>Fringilla coelebs</i>	Green		1	0	1	0	Foraging on site
Chiffchaff <i>Phylloscopus collybita</i>	Green		2	0	4	5	Probable breeding - 1 territory
Coot <i>Fulica atra</i>	Green		0	0	3	1	Probable breeding - 1 territory
Feral pigeon <i>Columba livia</i>	Green		1	0	0	1	Flying over the site
Goldfinch <i>Carduelis carduelis</i>	Green		4	2	2	2	Probable breeding – 2 territories
Great tit <i>Parus major</i>	Green		4	0	5	1	Probable breeding - 1 territory
Great spotted woodpecker <i>Dendrocopus major</i>	Green		2	0	0	0	Probable breeding - 1 territory
Jackdaw <i>Corvus monedula</i>	Green		0	0	1	1	Foraging within the site
Jay <i>Garrulus glandarius</i>	Green		1	0	3	0	Foraging within the site
Long-tailed tit <i>Aegithalos caudatus</i>	Green		5	4	5	4	Possible breeding - 2 territories

Species	BoCC	S. 41	V1	V2	V3	V4	Status
Magpie <i>Pica pica</i>	Green		6	14	3	0	Possible breeding – 1 territory
Pied wagtail <i>Motacilla alba</i>	Green		0	0	0	1	Foraging on the site
Reed warbler <i>Acrocephalus scirpaceus</i>	Green		0	0	0	1	Foraging on the site.
Robin <i>Erithacus rubecula</i>	Green		3	1	8	5	Probable breeding - 2 territories
Swallow <i>Hirundo rustica</i>	Green		2	2	1	0	Pairs in housing adjacent to site; foraging within site
Whitethroat <i>Sylvia communis</i>	Green		3	0	0	0	Possible breeding - 1 territory
Woodpigeon <i>Columba palumbus</i>	Green		1	1	0	0	Foraging on the site
Wren <i>Troglodytes troglodytes</i>	Green		5	1	5	3	Probable breeding - 2 territories
Little owl <i>Athene noctua</i>	Introduced		1	0	0	0	Hunting over the site
Pheasant <i>Phasianus colchicus</i>	Introduced		1	0	0	1	Possible breeding - 1 territory

I Introduced

N/A Not assessed

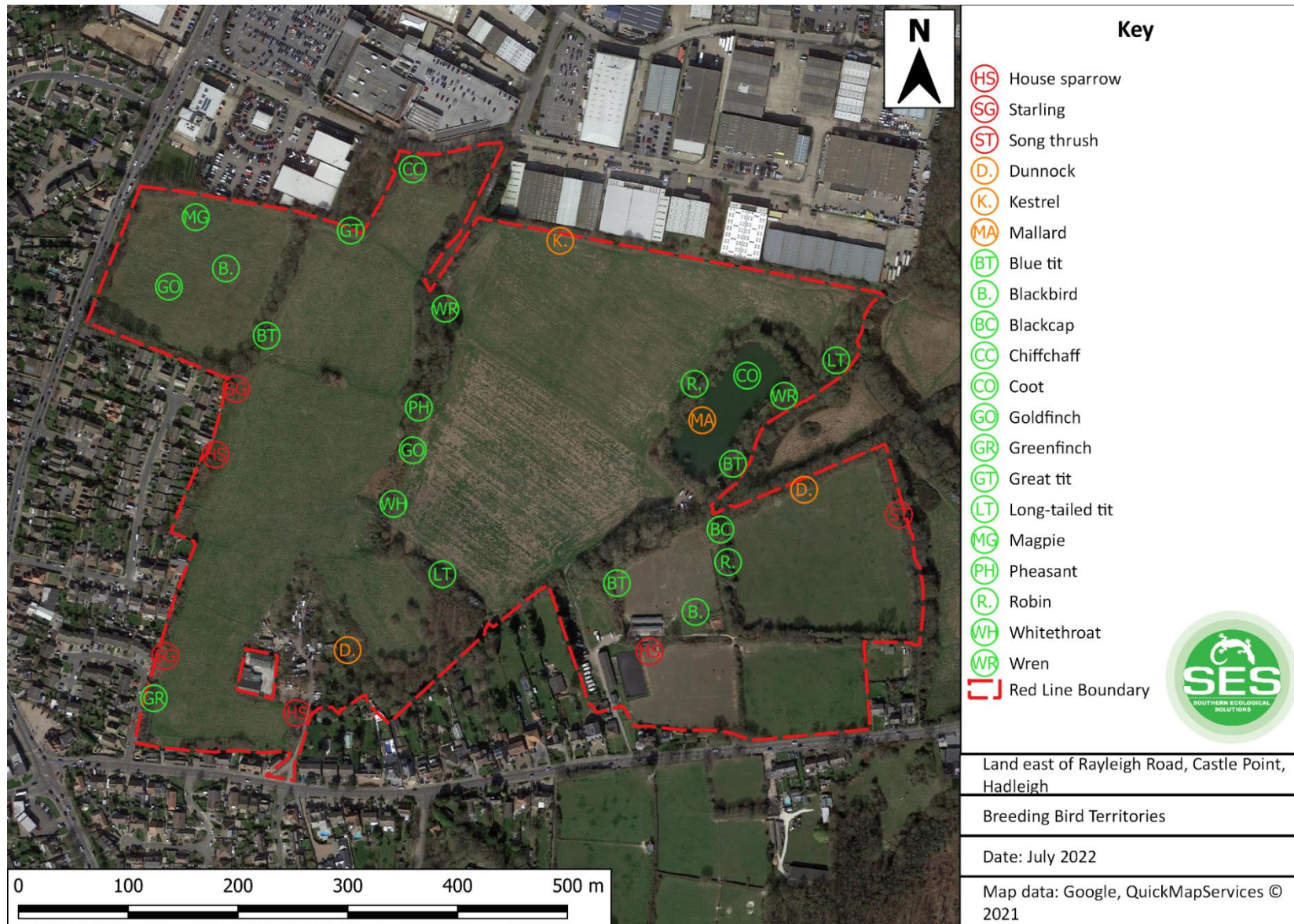
Red rows are BOCC red-list, Amber rows are BoCC amber-list, Green rows are BoCC green-list, NA rows are non-native species.

BoCC: Birds of Conservation Concern as defined and listed in Eaton *et al.*, (2021)

Table A9.2: Summary of breeding bird survey visit dates and weather conditions.

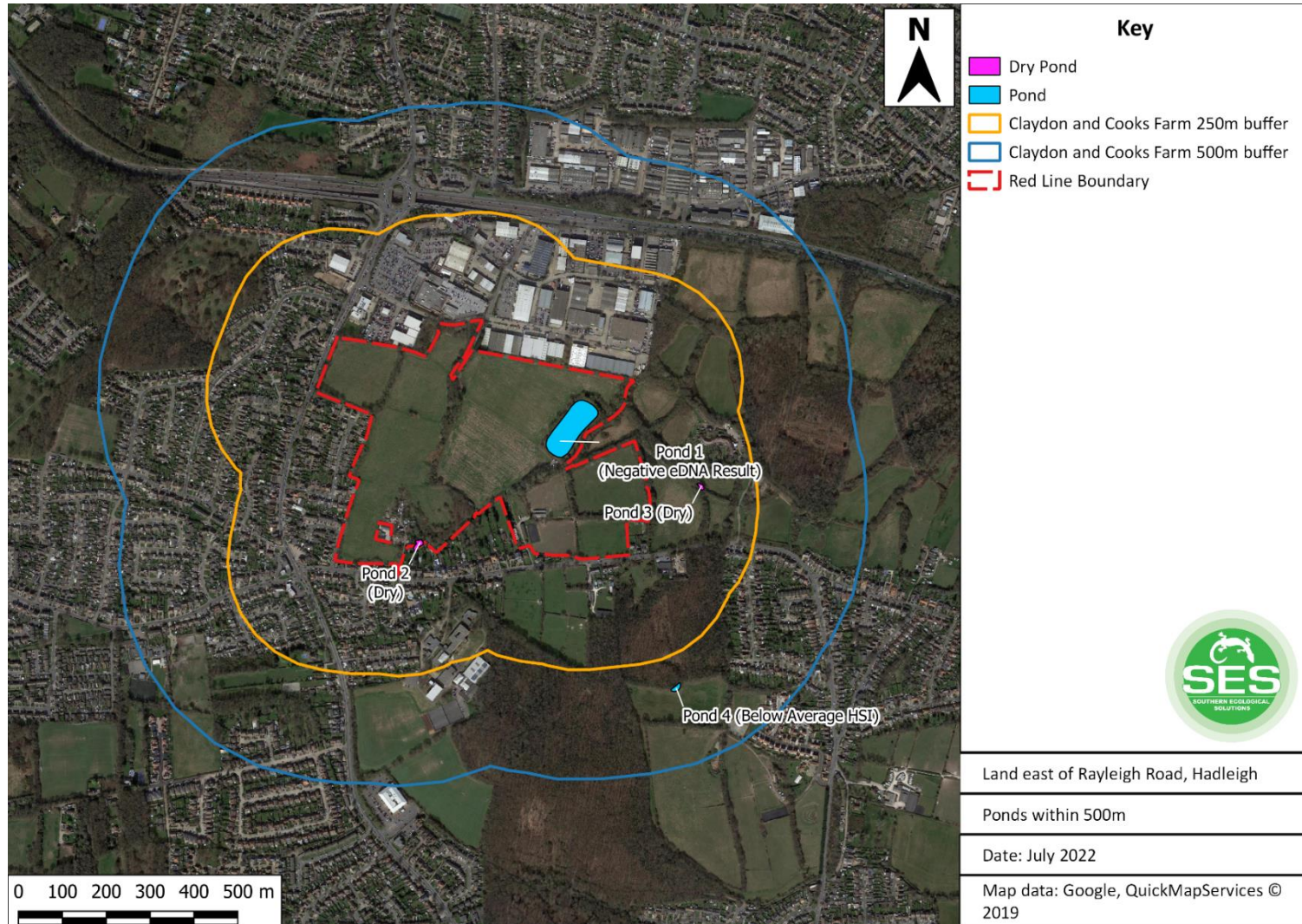
Visit	Date	Survey Conditions
1	17/06/2020	Good: 18°C (average), no precipitation, 1 wind, cloud 3/8, good visibility.
2	02/07/2020	Good: 18°C (average), no precipitation, 2 wind, cloud 5/8, good visibility.
3	13/04/2021	Good: 3°C (average), no precipitation, 1 wind, cloud 2/8, good visibility.
4	17/05/2021	Good: 11°C (average), no precipitation, 2 wind, cloud 8/8, good visibility.

Appendix 8b – Breeding Bird Survey Territory Map



Appendix 9: Great Crested Newt Survey Results

Appendix 9a – Locations of Ponds within 500m of the Site



Appendix 9b – HSI Survey Results

Table A9.1: Detailed HSI survey results.

Pond no.	1		4	
	Result	Score	Result	Score
1-Location	Zone A	1	A	1
2-Pond area	6720m ²	0.8	140m ²	0.3
3- Permanence	Never Dries	0.9	Dries Annually	0.1
4-Water quality	Good	1	Good	1
5- Shade	80%	0.6	100%	0.2
6- Waterfowl	Minor	0.67	Absent	1
7- Fish	Major	0	Absent	1
8- Pond count	4	0.72	5	0.75
9- Terrestrial habitat	Good	1	Good	1
10- Macrophytes	20%	0.5	10%	0.4
HSI Score	0.50		0.53	
Pond suitability	Below Average		Below Average	
Distance from the site	onsite		310m south	

Appendix 9c – eDNA Survey Results (June 2020)

Pond 1

Client: Josey Travell,
Southern Ecological Solutions



ADAS
Spring Lodge
172 Chester Road
Helsby
WAS 04N

Tel: 01159 518747
Email: Helen.Rees@adas.co.uk

www.adas.co.uk

Sample ID: 2020-1723 Condition on Receipt: Low Sediment Volume: Passed
Client Identifier: Pond 1 Rayleigh Road Description: pond water samples in preservative
Date of Receipt: 02/07/2020 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	2 of 2	Real Time PCR	16/07/2020
Degradation Control [‡]	Within Limits	Real Time PCR	16/07/2020
Great Crested Newt [§]	0 of 12 (GCN negative)	Real Time PCR	16/07/2020
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁴ ng/μL) [¶]	4 of 4	Real Time PCR	As above for GCN

Report Prepared by: Dr Helen Rees Report Issued by: Dr Ben Maddison

Signed:  Signed: 

Position: Director: Biotechnology Position: MD: Biotechnology

Date of preparation: 16/07/2020 Date of issue: 16/07/2020

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

[†] If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.

[‡] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

[§] No degradation is expected within time frame of kit preparation, sample collection and analysis.

[¶] Additional positive controls (10¹, 10², 10³ ng/μL) are also routinely run, results not shown here.

Pond 4

Client: Darren Denmede,
Southern Ecological Solutions



ADAS
Spring Lodge
172 Chiswick Road
Helsby
WAS 0AN



Tel: 01159 518747
Email: Helen.Rees@adas.co.uk

www.adas.co.uk

Sample ID: 2020-1356 Condition on Receipt: High Sediment Volume: Passed
Client Identifier: Pond 4 Raleigh Road Description: pond water samples in preservative
Date of Receipt: 02/07/2020 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control†	0 of 2	Real Time PCR	16/07/2020
Degradation Control‡	Evidence of degradation or residual inhibition	Real Time PCR	16/07/2020
Great Crested Newt*	Indeterminate	Real Time PCR	16/07/2020
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁴ ng/µL)*	4 of 4	Real Time PCR	As above for GCN

Report Prepared by: Dr Helen Rees Report Issued by: Dr Ben Maddison

Signed:  Signed: 

Position: Director: Biotechnology Position: MD: Biotechnology

Date of preparation: 16/07/2020 Date of issue: 16/07/2020

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 3 Technical Advice Note) published by DEFRA and adopted by Natural England.

** If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.*

† Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

‡ No degradation is expected within time frame of kit preparation, sample collection and analysis.

** Additional positive controls (10⁵, 10⁶, 10⁸ ng/µL) are also routinely run, results not shown here.*

Appendix 9d – eDNA Survey Results (April 2021)

Pond 4

Client: Laura Bennett,
SES Eco



ADAS
Spring Lodge
172 Chester Road
Helsby
WA6 0AR

Tel: 01159 516747
Email: Helen.Rees@adas.co.uk

www.adas.uk

Sample ID: ADAS-1282 Condition on Receipt: Low Sediment Volume: Passed
Client Identifier: Pond 4 Hadleigh Description: pond water samples in preservative
Date of Receipt: 29/04/2021 Material Tested: eDNA from pond water samples

Determinant	Result	Method	Date of Analysis
Inhibition Control [†]	0 of 2	Real Time PCR	11/05/2021
Degradation Control [§]	Within Limits	Real Time PCR	11/05/2021
Great Crested Newt*	0 of 12 (GCN negative)	Real Time PCR	11/05/2021
Negative PCR Control (Nuclease Free Water)	0 of 4	Real Time PCR	As above for GCN
Positive PCR Control (GCN DNA 10 ⁻⁴ ng/μL) [#]	4 of 4	Real Time PCR	As above for GCN

Report Prepared by: Dr Helen Rees Report Issued by: Dr Ben Maddison

Signed:

Signed:

Position: Director: Biotechnology Position: MD: Biotechnology

Date of preparation: 11/05/2021 Date of issue: 11/05/2021

eDNA analysis was carried out in accordance with the stipulated methodology found in the Technical Advice Note (WC1067 Appendix 5 Technical Advice Note) published by DEFRA and adopted by Natural England.

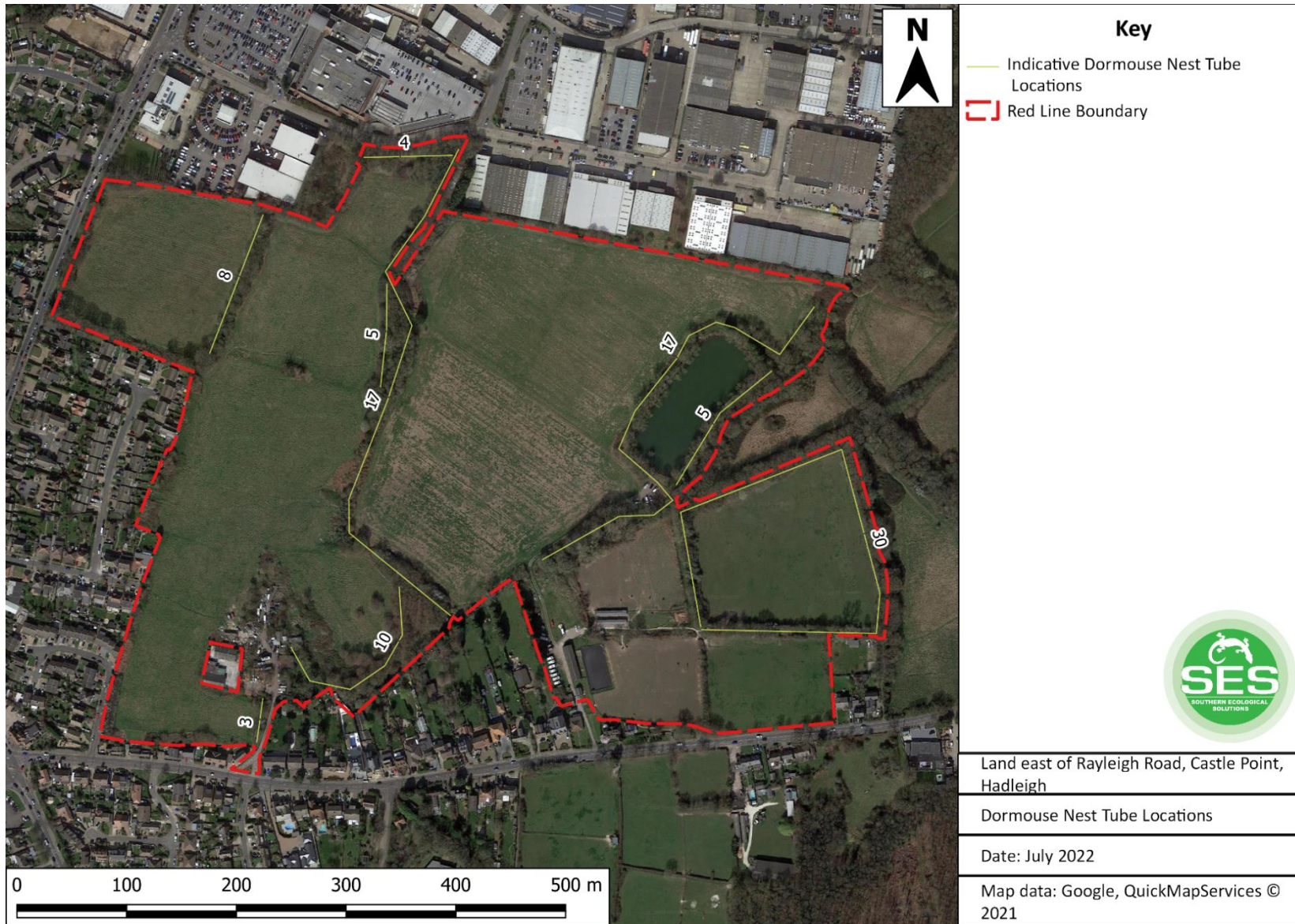
** If all PCR controls and extraction blanks give the expected results a sample is considered: negative for great crested newt if all of the replicates are negative; positive for great crested newt if one or more of the replicates are positive.*

[†] Recorded as the number of positive replicate reactions at expected C_t value. If the expected C_t value is not achieved, the sample is considered inhibited and is diluted as per the technical advice note prior to amplification with great crested newt primer and probes.

[§] No degradation is expected within time frame of kit preparation, sample collection and analysis.

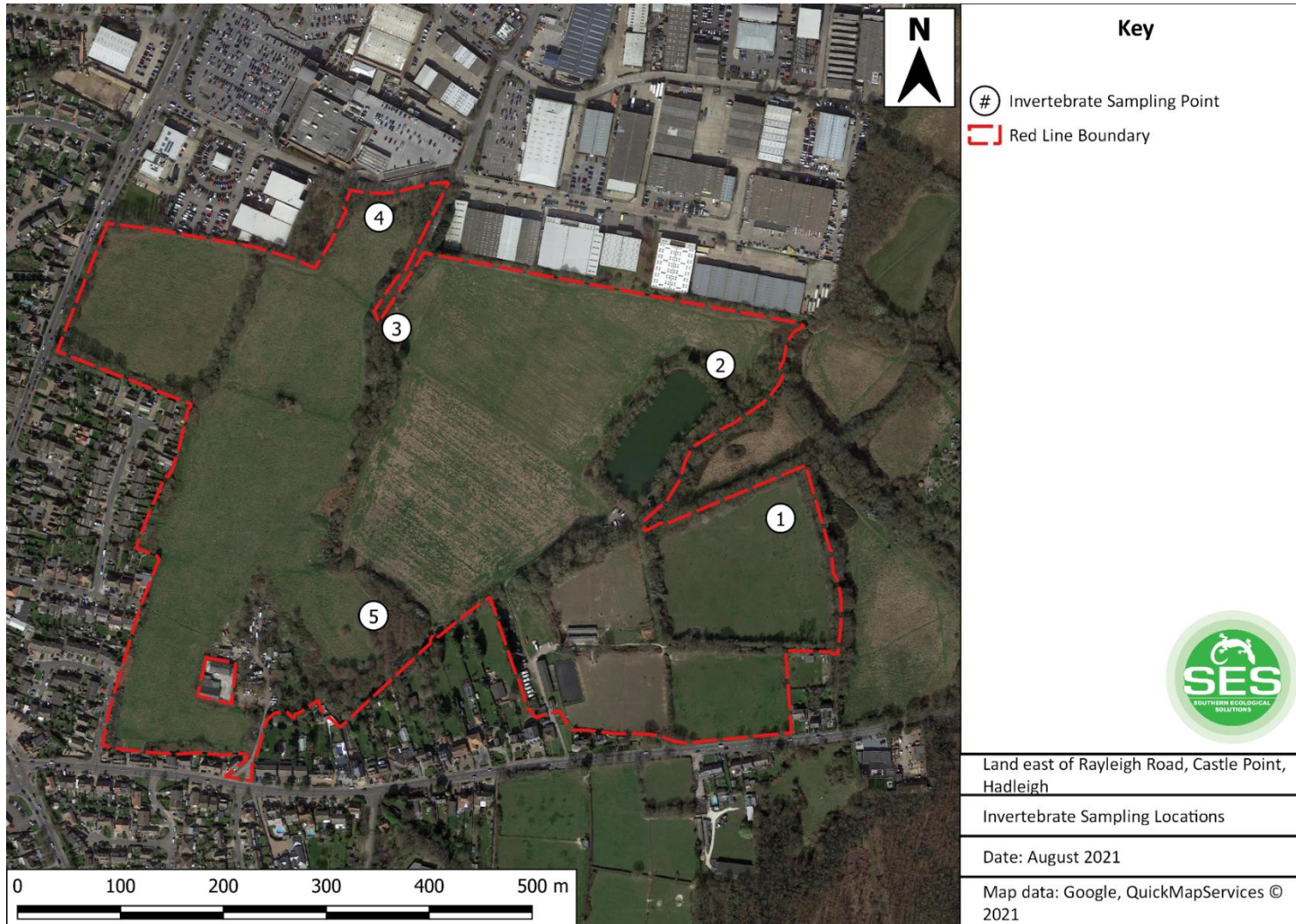
[#] Additional positive controls (10⁻¹, 10⁻², 10⁻³ ng/μL) are also routinely run, results not shown here.

Appendix 10. Dormouse Nest Tube Location Plan



Appendix 11: Invertebrate Survey Results

Appendix 11a – Locations of Invertebrate Sampling Stations



Appendix 11b – Combined survey information from 2020 and 2021

Table A12.1: Detailed species survey data.

Higher taxon	Family	Species	Sampling stations				
			1	2	3	4	5
Mollusca	Discidae	<i>Discus rotundatus</i>					X
Mollusca	Helicidae	<i>Cornu aspersum</i>					X
Mollusca	Helicidae	<i>Monacha cantiana</i>					X
Mollusca	Zonitidae	<i>Oxychilus allarius</i>				X	X
Molluscs	Helicidae	<i>Cepaea hortensis</i>					X
Isopoda	Armadillidae	<i>Armadillidium vulgare</i>					X
Isopoda	Oniscidae	<i>Oniscus asellus</i>					X
Isopoda	Philosciidae	<i>Philoscia muscorum</i>					X
Isopoda	Platyarthridae	<i>Platyarthrus hoffmannseggii</i>			X		
Isopoda	Porcellionidae	<i>Porcellio scaber</i>					X
Arachnida – Araneae	Araneidae	<i>Araneus diadematus</i>	X	X			X
Arachnida – Araneae	Araneidae	<i>Araniella cucurbitina</i>					X
Arachnida – Araneae	Clubionidae	<i>Cheiracanthium erraticum</i>		X			
Arachnida – Araneae	Clubionidae	<i>Clubiona comta</i>				X	
Arachnida – Araneae	Gnaphosidae	<i>Zelotes latreillei</i>				X	
Arachnida – Araneae	Linyphiidae	<i>Diplostyla concolor</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Dismodicus bifrons</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Erigone atra</i>			X		
Arachnida – Araneae	Linyphiidae	<i>Erigone dentipalpis</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Lepthyphantes tenuis</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Lepthyphantes zimmermanni</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Linyphia hortensis</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Neriene peltata</i>		X			
Arachnida – Araneae	Linyphiidae	<i>Tenuiphantes mengei</i>					X
Arachnida – Araneae	Linyphiidae	<i>Tenuiphantes tenuis</i>		X			
Arachnida – Araneae	Lycosidae	<i>Alopecosa pulverulenta</i>					X
Arachnida – Araneae	Lycosidae	<i>Trochosa terricola</i>		X			
Arachnida – Araneae	Nemastomatidae	<i>Nemastoma bimaculatum</i>			X		
Arachnida – Araneae	Phalangiidae	<i>Mitopus morio</i>		X			
Arachnida – Araneae	Phalangiidae	<i>Opilio saxatilis</i>					X
Arachnida – Araneae	Phalangiidae	<i>Phalangium opilio</i>					X
Arachnida – Araneae	Phalangiidae	<i>Platybunus triangularis</i>		X			
Arachnida – Araneae	Philodromidae	<i>Philodromus aureolus</i>					X
Arachnida – Araneae	Philodromidae	<i>Tibellus oblongus</i>		X			
Arachnida – Araneae	Pisauridae	<i>Pisaura mirabilis</i>			X		
Arachnida – Araneae	Salticidae	<i>Euophrys frontalis</i>					X
Arachnida – Araneae	Salticidae	<i>Heliophanus cupreus</i>			X		
Arachnida – Araneae	Salticidae	<i>Heliophanus flavipes</i>	X				
Arachnida – Araneae	Tetragnathidae	<i>Metellina mengei</i>			X		
Arachnida – Araneae	Theridiidae	<i>Enoplognatha ovata</i>	X			X	X
Arachnida – Araneae	Thomisidae	<i>Xysticus cristatus</i>					X
Arachnida – Araneae	Zoridae	<i>Zora spinimana</i>		X			
Coleoptera	Anthicidae	<i>Anthicus antherinus</i>				X	
Coleoptera	Apidae	<i>Anthophora plumipes</i>					X
Coleoptera	Apionidae	<i>Ceratapion gibbirostre</i>				X	
Coleoptera	Apionidae	<i>Ceratapion onopordi</i>				X	
Coleoptera	Apionidae	<i>Eutrichapion ervi</i>		X			

Higher taxon	Family	Species	Sampling stations				
			1	2	3	4	5
Coleoptera	Apionidae	<i>Ischnopterapion loti</i>					X
Coleoptera	Apionidae	<i>Protapion apricans</i>					X
Coleoptera	Cantharidae	<i>Cantharis nigra</i>				X	
Coleoptera	Cantharidae	<i>Rhagonycha fulva</i>		X			X
Coleoptera	Carabidae	<i>Amara convexior</i>					X
Coleoptera	Carabidae	<i>Badister bullatus</i>		X			
Coleoptera	Carabidae	<i>Demetrias atricapillus</i>		X			
Coleoptera	Carabidae	<i>Harpalus affinis</i>		X			
Coleoptera	Carabidae	<i>Harpalus rufipes</i>					X
Coleoptera	Carabidae	<i>Microlestes maurus</i>					X
Coleoptera	Carabidae	<i>Nebria brevicollis</i>		X			
Coleoptera	Carabidae	<i>Notiophilus biguttatus</i>	X	X			X
Coleoptera	Carabidae	<i>Paradromius linearis</i>				X	
Coleoptera	Carabidae	<i>Philorhizus melanocephalus</i>				X	
Coleoptera	Carabidae	<i>Pterostichus madidus</i>		X			
Coleoptera	Carabidae	<i>Pterostichus melanarius</i>		X			
Coleoptera	Carabidae	<i>Pterostichus nigrita</i>			X		
Coleoptera	Cerambycidae	<i>Agapanthia villosoviridescens</i>		X			
Coleoptera	Cerambycidae	<i>Clytus arietis</i>				X	
Coleoptera	Chrysomelidae	<i>Altica lythri</i>				X	
Coleoptera	Chrysomelidae	<i>Bruchus loti</i>	X				X
Coleoptera	Chrysomelidae	<i>Bruchus rufipes</i>					X
Coleoptera	Chrysomelidae	<i>Cassida rubiginosa</i>					X
Coleoptera	Chrysomelidae	<i>Longitarsus dorsalis</i>				X	
Coleoptera	Chrysomelidae	<i>Longitarsus pratensis</i>				X	
Coleoptera	Chrysomelidae	<i>Phaedon tumidulus</i>					X
Coleoptera	Chrysomelidae	<i>Phyllotreta atra</i>		X			
Coleoptera	Chrysomelidae	<i>Sphaeroderma testaceum</i>					X
Coleoptera	Coccinellidae	<i>Adalia bipunctata</i>		X		X	X
Coleoptera	Coccinellidae	<i>Coccinella septempunctata</i>				X	
Coleoptera	Coccinellidae	<i>Coccinella undecimpunctata</i>				X	
Coleoptera	Coccinellidae	<i>Harmonia axyridis</i>					X
Coleoptera	Coccinellidae	<i>Rhyzobius litura</i>		X			
Coleoptera	Coccinellidae	<i>Subcoccinella vigintiquatuorpunctata</i>		X			
Coleoptera	Coccinellidae	<i>Tytthaspis sedecimpunctata</i>					X
Coleoptera	Curculionidae	<i>Ceutorhynchus erysimi</i>		X			
Coleoptera	Curculionidae	<i>Hypera nigrirostris</i>					X
Coleoptera	Curculionidae	<i>Hypera postica</i>			X		
Coleoptera	Curculionidae	<i>Phyllobius virideaeris</i>				X	
Coleoptera	Curculionidae	<i>Sitona hispidulus</i>				X	
Coleoptera	Curculionidae	<i>Sitona humeralis</i>			X		
Coleoptera	Curculionidae	<i>Sitona lineatus</i>				X	
Coleoptera	Curculionidae	<i>Sitona puncticollis</i>			X		
Coleoptera	Curculionidae	<i>Sitona suturalis</i>	X				
Coleoptera	Curculionidae	<i>Trichosirocalus troglodytes</i>					X
Coleoptera	Elateridae	<i>Agriotes lineatus</i>		X			X
Coleoptera	Elateridae	<i>Agriotes sputator</i>					X
Coleoptera	Elateridae	<i>Athous haemorrhoidalis</i>				X	X
Coleoptera	Elateridae	<i>Kibunea minuta</i>		X			
Coleoptera	Latridiidae	<i>Corticarina minuta</i>			X		

Higher taxon	Family	Species	Sampling stations				
			1	2	3	4	5
Coleoptera	Malachiidae	<i>Malachius bipustulatus</i>		X		X	X
Coleoptera	Mordellidae	<i>Mordellistena neuwaldeggiana</i>		X			
Coleoptera	Oedemeridae	<i>Oedemera lurida</i>	X	X			
Coleoptera	Oedemeridae	<i>Oedemera nobilis</i>	X	X	X	X	X
Coleoptera	Silphidae	<i>Nicrophorus vespillo</i>					X
Coleoptera	Silphidae	<i>Silpha atrata</i>		X			
Coleoptera	Staphylinidae	<i>Drusilla canaliculata</i>		X			
Coleoptera	Staphylinidae	<i>Gyrophypnus fracticornis</i>		X			
Coleoptera	Staphylinidae	<i>Ischnosoma splendidum</i>					X
Coleoptera	Staphylinidae	<i>Metopsia clypeata</i>					X
Coleoptera	Staphylinidae	<i>Mocyta fungi</i>		X			
Coleoptera	Staphylinidae	<i>Quedius semiobscurus</i>		X			
Coleoptera	Staphylinidae	<i>Sepedophilus nigripennis</i>				X	
Coleoptera	Staphylinidae	<i>Stenus aceris</i>			X		
Coleoptera	Staphylinidae	<i>Stenus ossium</i>		X			
Coleoptera	Staphylinidae	<i>Tachyporus atriceps</i>		X			
Coleoptera	Staphylinidae	<i>Tachyporus dispar</i>			X		
Coleoptera	Staphylinidae	<i>Tachyporus hypnorum</i>				X	
Coleoptera	Staphylinidae	<i>Tachyporus pusillus</i>			X		
Coleoptera	Staphylinidae	<i>Tachyporus solutus</i>			X		
Coleoptera	Tenebrionidae	<i>Lagria hirta</i>	X			X	
Dermaptera	Forficulidae	<i>Forficula auricularia</i>					X
Diptera	Asilidae	<i>Dioctria atricapilla</i>	X	X			X
Diptera	Asilidae	<i>Dioctria baumhaueri</i>					X
Diptera	Asilidae	<i>Dioctria rufipes</i>		X			X
Diptera	Asilidae	<i>Leptogaster cylindrica</i>	X			X	
Diptera	Bibionidae	<i>Bibio marci</i>	X	X			
Diptera	Bibionidae	<i>Dilophus febrilis</i>	X				
Diptera	Bombyliidae	<i>Bombylius major</i>		X			
Diptera	Calliphoridae	<i>Calliphora vicina</i>					X
Diptera	Calliphoridae	<i>Lucilia caesar</i>					X
Diptera	Chloropidae	<i>Elachiptera cornuta</i>				X	
Diptera	Conopidae	<i>Conops quadrifasciatus</i>				X	
Diptera	Conopidae	<i>Myopa buccata</i>					X
Diptera	Conopidae	<i>Sicus ferrugineus</i>	X	X			
Diptera	Dolichopodidae	<i>Dolichopus festivus</i>		X			
Diptera	Dolichopodidae	<i>Dolichopus griseipennis</i>		X			
Diptera	Dolichopodidae	<i>Hercostomus cupreus</i>		X			
Diptera	Empididae	<i>Empis aestiva</i>		X			
Diptera	Empididae	<i>Empis livida</i>	X	X			
Diptera	Empididae	<i>Empis nigripes</i>		X			
Diptera	Hybotidae	<i>Platypalpus annulipes</i>					X
Diptera	Lauxaniidae	<i>Tricholauxania praeusta</i>		X			
Diptera	Lonchopteridae	<i>Lonchoptera lutea</i>			X		
Diptera	Muscidae	<i>Coenosia tigrina</i>					X
Diptera	Muscidae	<i>Helina evecta</i>					X
Diptera	Rhagionidae	<i>Chrysopilus cristatus</i>		X			
Diptera	Rhagionidae	<i>Rhagio scolopaceus</i>		X			
Diptera	Rhinophoridae	<i>Paykullia maculata</i>				X	
Diptera	Sarcophagidae	<i>Brachicoma devia</i>					X
Diptera	Scathophagidae	<i>Scathophaga stercoraria</i>			X		

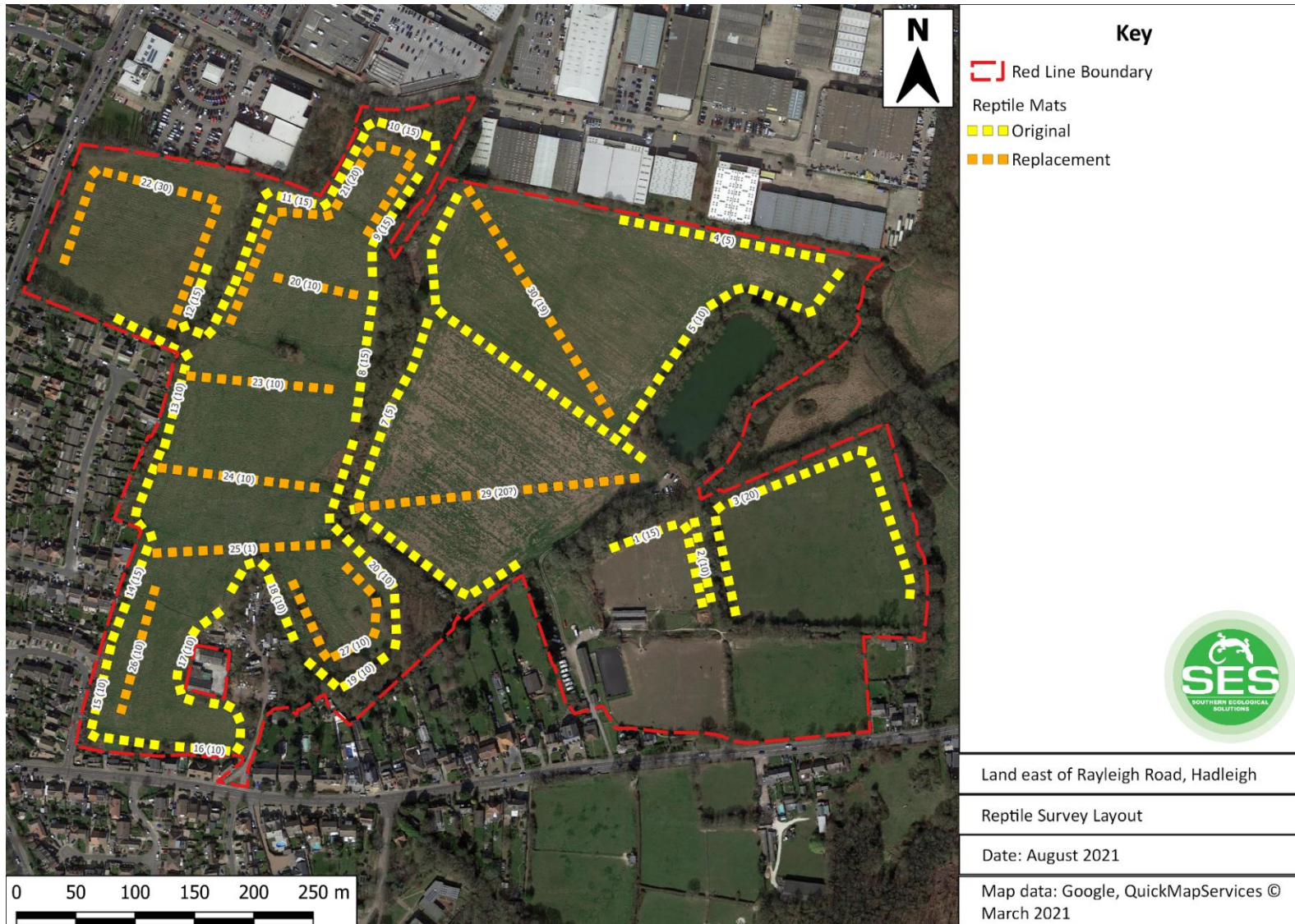
Higher taxon	Family	Species	Sampling stations					
			1	2	3	4	5	
Diptera	Sepsidae	<i>Sepsis fulgens</i>						X
Diptera	Stratiomyidae	<i>Beris chalybata</i>		X				
Diptera	Stratiomyidae	<i>Chloromyia formosa</i>	X	X				
Diptera	Stratiomyidae	<i>Chorisops tibialis</i>		X				
Diptera	Stratiomyidae	<i>Sargus bipunctatus</i>		X				
Diptera	Syrphidae	<i>Helophilus pendulus</i>			X			
Diptera	Syrphidae	<i>Cheilosia albitarsis</i>					X	X
Diptera	Syrphidae	<i>Cheilosia bergenstammi</i>					X	
Diptera	Syrphidae	<i>Cheilosia variabilis</i>		X				
Diptera	Syrphidae	<i>Epistrophe eligans</i>		X				
Diptera	Syrphidae	<i>Episyrphus balteatus</i>		X			X	X
Diptera	Syrphidae	<i>Eristalinus sepulchralis</i>		X				
Diptera	Syrphidae	<i>Eristalis arbustorum</i>	X		X			X
Diptera	Syrphidae	<i>Eristalis intricarius</i>						X
Diptera	Syrphidae	<i>Eristalis nemorum</i>			X			X
Diptera	Syrphidae	<i>Eristalis pertinax</i>		X				
Diptera	Syrphidae	<i>Eristalis tenax</i>		X			X	X
Diptera	Syrphidae	<i>Eupeodes luniger</i>					X	
Diptera	Syrphidae	<i>Melanostoma mellinum</i>					X	
Diptera	Syrphidae	<i>Merodon equestris</i>	X					
Diptera	Syrphidae	<i>Neosciasca tenur</i>					X	X
Diptera	Syrphidae	<i>Platycheirus albimanus</i>		X				
Diptera	Syrphidae	<i>Platycheirus clypeatus</i>	X	X				
Diptera	Syrphidae	<i>Platycheirus manicatus</i>			X			
Diptera	Syrphidae	<i>Sphaerophoria scripta</i>		X				
Diptera	Syrphidae	<i>Syritta pipiens</i>	X	X	X	X	X	X
Diptera	Syrphidae	<i>Syrphus ribesii</i>		X				
Diptera	Syrphidae	<i>Volucella pellucens</i>		X				
Diptera	Syrphidae	<i>Volucella zonaria</i>						X
Diptera	Tabanidae	<i>Haematopota pluvialis</i>		X				
Diptera	Tephritidae	<i>Tephritis cometa</i>		X				
Diptera	Tephritidae	<i>Tephritis formosa</i>		X				
Diptera	Tephritidae	<i>Urophora cardui</i>	X	X				
Diptera	Tipulidae	<i>Limonia nubeculosa</i>		X				X
Diptera	Tipulidae	<i>Nephrotoma flavescens</i>						X
Diptera	Tipulidae	<i>Tipula fascipennis</i>		X				
Diptera	Tipulidae	<i>Tipula oleracea</i>		X				
Hemiptera - Heteroptera	Acanthosomatidae	<i>Elasmucha grisea</i>	X				X	X
Hemiptera - Heteroptera	Coreidae	<i>Coriomeris denticulatus</i>	X					
Hemiptera - Heteroptera	Lygaeidae	<i>Cymus melanocephalus</i>					X	
Hemiptera - Heteroptera	Lygaeidae	<i>Heterogaster urticae</i>			X			X
Hemiptera - Heteroptera	Miridae	<i>Apolygus lucorum</i>						X
Hemiptera - Heteroptera	Miridae	<i>Calocoris norvegicus</i>		X				X
Hemiptera - Heteroptera	Miridae	<i>Capsus ater</i>			X			X
Hemiptera - Heteroptera	Miridae	<i>Closterotomus norwegicus</i>	X	X	X	X	X	X
Hemiptera - Heteroptera	Miridae	<i>Deraeocoris ruber</i>					X	
Hemiptera - Heteroptera	Miridae	<i>Leptopterna dolabrata</i>		X				
Hemiptera - Heteroptera	Miridae	<i>Liocoris tripustulatus</i>					X	
Hemiptera - Heteroptera	Miridae	<i>Lygocoris pabulinus</i>					X	X
Hemiptera - Heteroptera	Miridae	<i>Lygus rugulipennis</i>					X	X
Hemiptera - Heteroptera	Miridae	<i>Notostira elongata</i>						X

Higher taxon	Family	Species	Sampling stations				
			1	2	3	4	5
Hemiptera - Heteroptera	Miridae	<i>Pithanus maerkelii</i>	X	X			
Hemiptera - Heteroptera	Miridae	<i>Stenodema calcarata</i>				X	
Hemiptera - Heteroptera	Nabidae	<i>Nabis flavomarginatus</i>	X				
Hemiptera - Heteroptera	Pentatomidae	<i>Dolycoris baccarum</i>				X	X
Hemiptera - Heteroptera	Pentatomidae	<i>Eurydema oleracea</i>	X				
Hemiptera - Heteroptera	Pentatomidae	<i>Palomena prasina</i>		X			X
Hemiptera - Heteroptera	Pentatomidae	<i>Pentatoma rufipes</i>				X	X
Hemiptera - Heteroptera	Pentatomidae	<i>Podops inuncta</i>	X				
Hemiptera - Heteroptera	Pentatomidae	<i>Troilus luridus</i>		X			
Hemiptera: Auchenorrhyncha	Aphrophoridae	<i>Aphrophora alni</i>		X			
Hemiptera: Auchenorrhyncha	Aphrophoridae	<i>Neophilaenus lineatus</i>		X			
Hemiptera: Auchenorrhyncha	Aphrophoridae	<i>Philaenus spumarius</i>		X			
Hemiptera: Auchenorrhyncha	Cicadellidae	<i>Cicadella viridis</i>		X			
Hemiptera: Auchenorrhyncha	Cicadellidae	<i>Eupteryx urticae</i>	X				
Hemiptera: Auchenorrhyncha	Cicadellidae	<i>Euscelis incisus</i>			X		
Hemiptera: Auchenorrhyncha	Cicadellidae	<i>Zyginidia scutellaris</i>		X			
Hemiptera: Auchenorrhyncha	Delphacidae	<i>Javesella pellucida</i>		X			
Hemiptera: Auchenorrhyncha	Delphacidae	<i>Stenocranus minutus</i>				X	
Hemiptera: Heteroptera	Acanthosomatidae	<i>Acanthosoma haemorrhoidale</i>					X
Hemiptera: Heteroptera	Anthocoridae	<i>Anthocoris confusus</i>				X	
Hemiptera: Heteroptera	Coreidae	<i>Coreus marginatus</i>		X			
Hemiptera: Heteroptera	Lygaeidae	<i>Nysius huttoni</i>				X	
Hemiptera: Heteroptera	Miridae	<i>Deraeocoris lutescens</i>		X			
Hemiptera: Heteroptera	Miridae	<i>Stenodema laevigata</i>				X	
Hemiptera: Heteroptera	Nabidae	<i>Himacerus mirmicoides</i>					X
Hemiptera: Heteroptera	Nabidae	<i>Nabis ferus</i>		X		X	
Hemiptera: Heteroptera	Pentatomidae	<i>Aelia acuminata</i>	X				
Hemiptera: Heteroptera	Rhopalidae	<i>Corizus hyoscyami</i>				X	
Hemiptera: Heteroptera	Rhopalidae	<i>Rhopalus subrufus</i>		X			
Hemiptera: Heteroptera	Tingidae	<i>Tingis cardui</i>		X			X
Hymenoptera	Andrenidae	<i>Andrena dorsata</i>				X	
Hymenoptera	Andrenidae	<i>Andrena flavipes</i>					X
Hymenoptera	Andrenidae	<i>Andrena fulva</i>					X
Hymenoptera	Andrenidae	<i>Andrena praecox</i>					X
Hymenoptera	Apidae	<i>Bombus hortorum</i>			X	X	
Hymenoptera	Apidae	<i>Bombus hypnorum</i>		X			X
Hymenoptera	Apidae	<i>Bombus lapidarius</i>	X				
Hymenoptera	Apidae	<i>Bombus pascuorum</i>	X	X	X	X	
Hymenoptera	Apidae	<i>Bombus sylvarum</i>	X				
Hymenoptera	Apidae	<i>Bombus terrestris</i>	X				
Hymenoptera	Apidae	<i>Colletes similis</i>				X	
Hymenoptera	Apidae	<i>Halictus rubicundus</i>				X	
Hymenoptera	Apidae	<i>Halictus tumulorum</i>				X	X
Hymenoptera	Apidae	<i>Nomada goodeniana</i>	X				
Hymenoptera	Chrysididae	<i>Chrysis ignita</i>				X	

Higher taxon	Family	Species	Sampling stations				
			1	2	3	4	5
Hymenoptera	Colletidae	<i>Hylaeus communis</i>		X			
Hymenoptera	Crabronidae	<i>Cerceris rybyensis</i>	X				
Hymenoptera	Crabronidae	<i>Ectemnius continuus</i>					X
Hymenoptera	Formicidae	<i>Formica fusca</i>				X	
Hymenoptera	Formicidae	<i>Lasius niger</i>	X		X		X
Hymenoptera	Formicidae	<i>Myrmica scabrinodis</i>		X			
Hymenoptera	Halictidae	<i>Lasioglossum leucopus</i>	X			X	
Hymenoptera	Halictinae	<i>Lasioglossum calceatum</i>	X				
Hymenoptera	Megachilidae	<i>Megachile versicolor</i>		X			X
Hymenoptera	Vespidae	<i>Gymnomerus laevipes</i>		X			
Hymenoptera	Vespidae	<i>Vespula vulgaris</i>	X			X	
Lepidoptera	Arctiidae	<i>Tyria jacobaeae</i>	X	X			X
Lepidoptera	Hesperiidae	<i>Thymelicus sylvestris</i>	X				X
Lepidoptera	Hesperiidae	<i>Ochlodes faunus</i>					X
Lepidoptera	Hesperiidae	<i>Thymelicus lineola</i>	X				X
Lepidoptera	Lepidoptera	<i>Aphantopus hyperantus</i>	X	X		X	X
Lepidoptera	Lycaenidae	<i>Polyommatus icarus</i>		X			
Lepidoptera	Lycaenidae	<i>Lycaena phlaeas</i>		X		X	X
Lepidoptera	Nymphalidae	<i>Aglais io</i>				X	
Lepidoptera	Nymphalidae	<i>Inachis io</i>	X				X
Lepidoptera	Nymphalidae	<i>Vanessa atalanta</i>		X			
Lepidoptera	Pieridae	<i>Colias croceus</i>	X				
Lepidoptera	Pieridae	<i>Pieris brassicae</i>			X		
Lepidoptera	Satyridae	<i>Coenonympha pamphilus</i>	X				X
Lepidoptera	Satyridae	<i>Maniola jurtina</i>	X	X			
Lepidoptera	Satyridae	<i>Melanargia galathea</i>					X
Lepidoptera	Satyridae	<i>Pararge aegeria</i>			X		
Lepidoptera	Satyridae	<i>Pyronia tithonus</i>		X			
Lepidoptera	Nymphalidae	<i>Polygonia c-album</i>	X				
Mecoptera	Panorpidae	<i>Panorpa communis</i>		X			
Neuroptera	Chrysopidae	<i>Hemerobius humulinus</i>		X			
Orthoptera	Acrididae	<i>Chorthippus brunneus</i>				X	X
Orthoptera	Acrididae	<i>Chorthippus parallelus</i>		X			
Orthoptera	Conocephalidae	<i>Conocephalus fuscus</i>		X			
Orthoptera	Meconematodae	<i>Meconema thalassinum</i>			X		
Orthoptera	Phaneropteridae	<i>Leptophyes punctatissima</i>					X
Orthoptera	Tetrigidae	<i>Tetrix undulata</i>		X			
Orthoptera	Tettigoniidae	<i>Metrioptera roeselii</i>		X			X

Appendix 12: Reptile Survey Results

Appendix 12a – Reptile Refugia Location Plan



Appendix 12b – Reptile Survey Results

Table A13.1: Detailed reptile survey results.

Visit No.	Date	Temperature (°C)	Cloud (%)	Wind (Beaufort)	Precipitation	Species	Location
1	22/07/2020	16	60	1	0	2 x female common lizard 5 x female slow worm	Grassland habitats – Field Compartments 1, 2 & 3.
2	29/07/2020	18	0	1	0	1 x male common lizard 3 x slow worm (1 female, 2 male)	Grassland habitats – Field Compartment 1
3	03/08/2020	18	0	0	0	2 x slow worm (1 male, 1 juvenile)	Grassland habitats – Field Compartments 1 & 2.
4	05/08/2020	18	0	1	0	2 x slow worm (1 male, 1 juvenile)	Grassland habitats - Field Compartment 1
5	17/08/2020	17	10	1	0	5 x slow worm (3 female, 2 male)	Grassland habitats - Field Compartments 1 & 2.
6	29/08/2020	15	80	0	0	2 x female common lizard	Grassland habitats - Field Compartment 1
7	23/09/2020	17	100	1	0	2 x female common lizard	Grassland habitats - Field Compartment 2

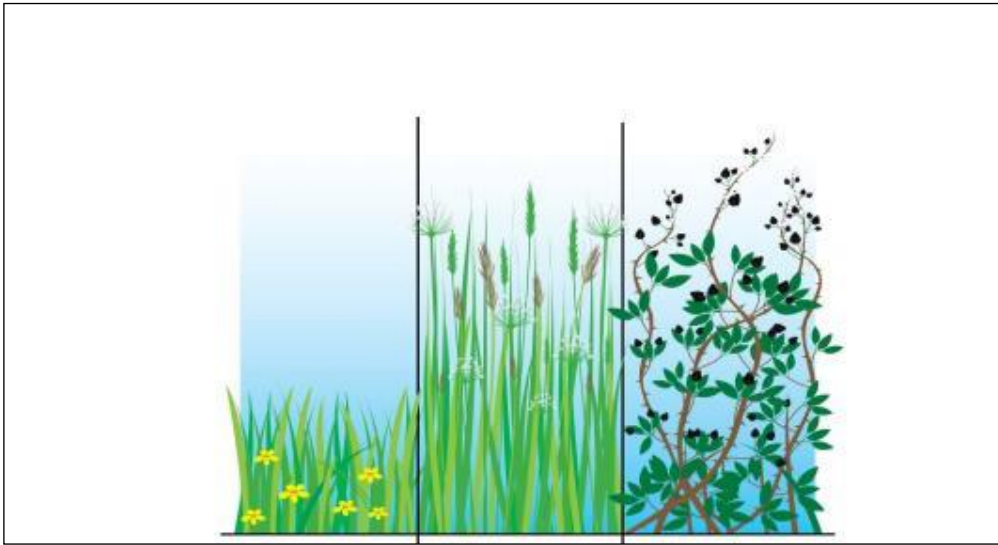
Appendix 13: Habitat Creation and Management Summary

Specific detail to be provided within the LEMP, below is a guide to establish feature habitats.

Scrub grassland

- The seed mix (EM10 or similar) will be sown in late spring or autumn
- The grassland will be mown regularly throughout the first year of establishment to a height of 40-60mm to control annual weeds and help maintain balance between faster growing grasses and slower developing wildflowers.
- Post-establishment, tussock grassland areas will be cut on a 3 year rotation, with no more than a third of the total habitat resource or two hectares (whichever is smaller) cut in any one year (Edgar, Foster & Baker 2010). This will ensure continuous provision of longer-sward grassland suitable for reptiles and attractive to invertebrate species. Cutting of tussock grassland areas will be undertaken during the winter period of reptile inactivity from November to February when individuals will be concentrated within scrub, roots and purpose-built hibernacula refuge habitats, to minimise risk of harm. Arisings will be collected and used to form compost piles located close to reptile hibernacula, to provide egg-laying site for grass snake in the spring.
- Natural features that develop such as ant hills will be retained and not destroyed as they create valuable micro habitats for animals and plants.
- This regime will create a biodiverse grass sward and in turn provide a valuable habitat for invertebrates, foraging bats, badgers and breeding and over wintering birds coupled with nesting habitat provided within the built development and biodiversity building which complement the provision of this habitat.
- Islands of species rich scrub will be planted with November – February. These islands will be mapped and a maximum extent of scrub encroachment will be established with managed with the aim of creating a gradient of scrub/ tall herbs to grassland of varied ages (see figure below). This will be achieved through 5 year rotational cuts (dictated by ground conditions). These cuts will be outside of nesting bird season March to August (inclusive)

Scrub grading to tall ruderal and grassland



Meadow grassland

- Wildflower grassland will be created using Emorsgate Seeds mixture EM3 (or similar suitable mix) with an increased proportion of yellow rattle versus the standard mix, as this mix is suitable where precise soil and site characteristics have not been established before sowing.
- Identified areas will be sown in September. Prior to sowing, any existing vegetation will be tightly cut, clippings will be removed, and the area scarified with a harrow to reveal bare soil. Spot spraying of all competitive weeds such as for thistle *Cirsium sp.* and ragwort *Jacobaea vulgaris* will also be undertaken before seed is sown.
- Yellow rattle *Rhinanthus minor* will be seeded along with the meadow mix as this is a parasite on and reduces grass; thus its inclusion in the planting mix should help to aid the establishment of wildflowers which may otherwise be outcompeted by fast growing grasses (English Nature/Wildlife Trusts, 1999). Yellow rattle will be sourced from a reputable supplier (see Supplier List on www.floralocale.org).
- In year 1 the grassland will be maintained to a height of 200mm between May and July. This will be above the height of the yellow rattle, allowing time for this species to set seed with the intention of achieving blanket coverage of the meadow within 5 years. In July, the grassland will be cut short to <20mm (short cuttings will dry out quickly so will not smother seedlings).
- In year 2, weeds which have grown back from the seed bank will be topped via a main cut to 150mm during mid-summer, with a final cut to <20mm undertaken in mid-October. The latter will replicate aftermath grazing as in traditional hay meadow management (English Nature/Wildlife Trusts, 1999). Arisings will be removed and used to form compost heaps adjacent to reptile hibernacula.
- In years 1 and 2 the wildflower meadow will be protected from disturbance by restricting access with temporary barrier fencing and signage.

- Year 3 will follow the same pattern as year 2. If the winter has been warm and dry, an additional spring cut to <20mm may be necessary to remove early growth that could swamp wildflowers later in the year; the need for this measure will be determined by the on-site ECoW.
- Throughout the first 3 years of establishment, the ECoW will undertake a yearly 'snagging' visit during late spring to highlight any challenges to successful establishment such as weed problems. Additional spot spraying/ pulling is likely to be required on an ongoing basis. Spot spraying is most effective and will be undertaken in April to early May. A detailed review will be carried out in year 4 and plans adjusted accordingly with the repetition of year 3 actions. If good coverage of yellow rattle has not been achieved across the wildflower meadow area, additional seeds will be sown in autumn of year 4 following the short October cut.
- Throughout and beyond establishment, timing of the main summer cut will be varied year on year to maximise the structural and species diversity of the sward, alternating between early cuts (in July) to later cuts (in August to September). A rotational cutting regime will be followed whereby no more than one third of the total wildflower meadow habitat resource is cut at any one time.
- It is predicted that wildflowers will spread into surrounding grassland as the habitat matures.

Wet grassland

Areas of wet grassland will be sown with EM8 / 4 (or similar). Management as per meadow grassland.

Broadleaved Woodland

- Native woody species will be planted explicitly not in straight lines with centres every 2-3m. Species will be native mimicking that of the lowland deciduous woodland onsite with the intention of creating a multi story woodland. A thorny shelter belt will be planted to provide protection from excess recreational pressure.
- Planting should occur between November – March, preferably in November after the first frost. A dibber should be used to plant bare root trees with suitable rabbit and deer browsing guards. Spot spraying of competitive vegetation should occur in the spring. Large scale failures should be replaced but failures on a smaller scale should be tolerated as this could provide more diverse, varied woodland.
- Woodland should be thinned on a 7 year rotation with dead wood.

Orchard

Orchard management

- An orchard will be planted with species of local provenance. Due to modern day farming methods and how we source fruit; orchards within the UK have been uneconomic and thus have suffered major declines since the 1950's. Orchards managed in a low intensity manner produce a rich biodiverse habitat.

Orchard Trees

- For orchard biodiversity to flourish a variety of trees of varying age are required. Fruit trees are generally, although not necessarily, short-lived trees compared to other hardwood species. This means that they

begin to produce veteran tree features such as hollow trunks, rot holes, split bark, tears, lightning strikes and sap runs relatively quickly. Because of the wide tree spacing in orchards compared to woodland, the dead and decaying wood is usually in open, sunny locations. These conditions create good habitat for insects and other invertebrate species which depend on decaying wood habitats.

Management Requirements

- As in parkland or wood pasture sites, careful management of the trees is necessary to maintain older individual ones. Dead and decaying branches should not be removed unless they interfere with necessary operations or are unsafe. Large cut branches, fallen dead wood or remains of old trees should be left on site. Planned replanting over time and adequate aftercare of new fruit trees is also required to ensure the long term future of the orchard.
- The abundance of climbers such as bramble and ivy should be controlled to allow plants and animals that need higher light levels or warmth to survive.
- Any tree health problems need to be assessed on a case by case basis. The application of chemicals to control pests and diseases should be minimised, and their use should be the exception rather than the rule.

Orchard Floor Management

- The grasses at the base of the orchard floor should be cut in late summer and again in the September/October. Grass arising should be removed rather than left in situ to prevent soil nutrients building up and therefore favouring competitive species at the expense of a varied sward. Further cuts may be needed should scrub come a problem. A network of mown paths should be maintained to allow people to enjoy and forage from the orchard.
- Many species of bumblebee, solitary bee and other insects may be present in an orchard. Wild bees play an important role during the blossom period when they help to pollinate the orchard trees. They are particularly helpful as they are active in colder conditions and forage for longer than honey bees. Many bee species use pollen and nectar from flowers in the orchard grassland and tall herb areas as well as fruit blossom on the trees. Members of the daisy family (such as ox-eye daisy *Leucanthemum vulgare*), hogweed *Heracleum sphondylium* and other umbellifers and legumes (clovers, vetches and trefoils) are particularly important food sources.

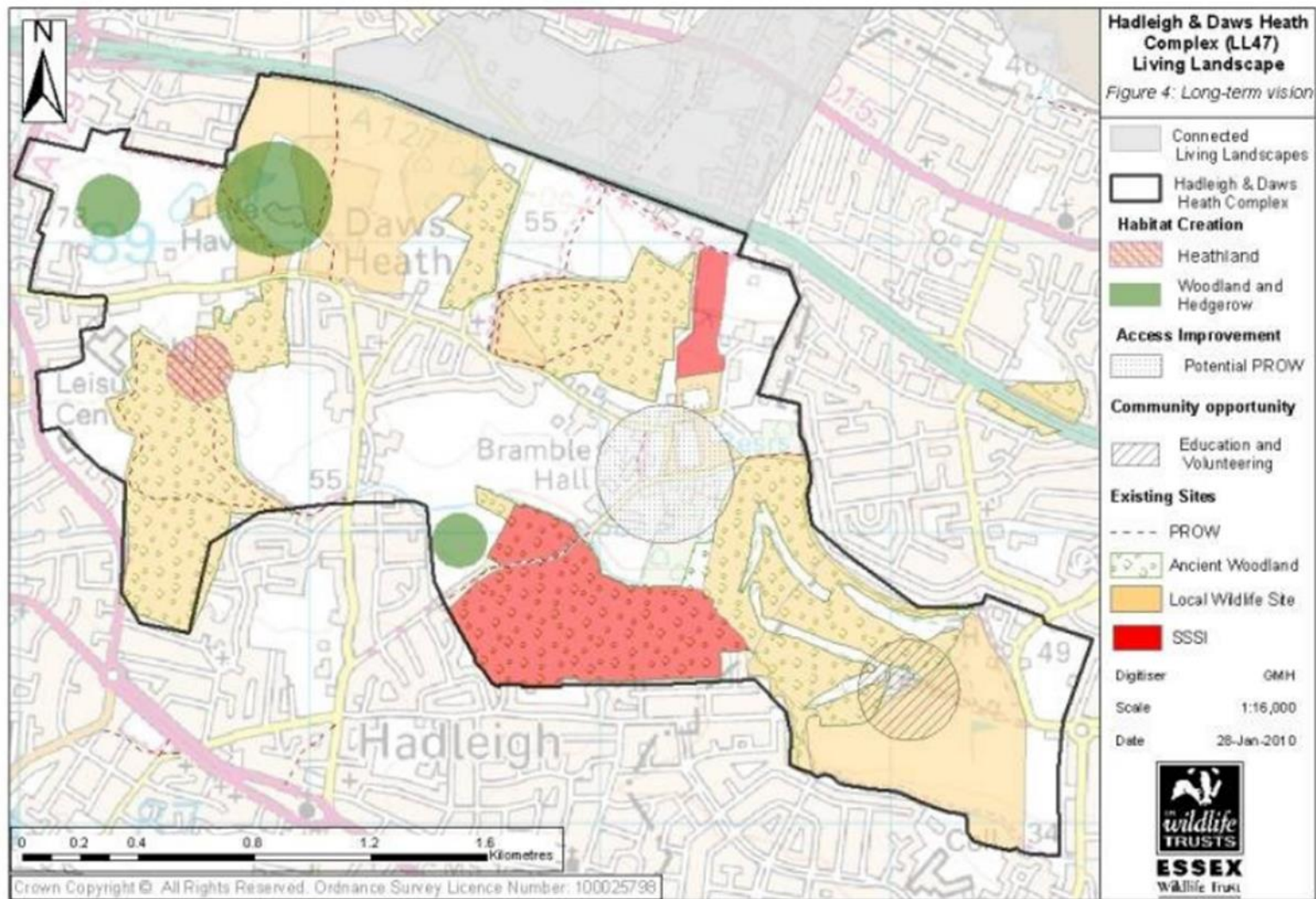
Hedgerows

- The site has been master planned to retain key hedgerows where possible. Existing hedgerows will be 'gapped up' where necessary to ensure that no gaps are presented larger than 10% of the hedgerow. Planting of whips with rabbit guards will be undertaken from November – February. Fruiting species will also be used to gap up hedgerows providing a valuable late season food source for many species. New hedgerows will be species rich (10 species including fruiting varieties see Appendix 5 for suggested planting list) with at least one standard tree per 50m. Retained and created hedgerows will be maintained in a low intensity manner meaning that cutting is carried out on 2-3 year rotation this will allow the development of hedge that is diverse in structure and prevent tall and woody 'leggy' hedges developing. This structure will allow a greater yield of fruit, nuts and seeds to be produced

which in turn will benefit many species including farmland birds. The transition between hedgerow and grassland habitats will have a buffer of 2m (minimum of 1m) to reduce 'edge' effect and also provide a greater habitat mosaic. This buffer will consist of grasses and tall ruderal vegetation.

- Rotational cutting should take place outside of bird nesting season, as a guide March – August. If this is not possible it may be appropriate on occasion to cut hedgerows within bird nesting season after an ecologist has declared the area in question to be free from nesting birds.

Appendix 14: Hadleigh and Daws Heath Complex Living Landscape Long-Term Vision



Appendix 15: Badger Survey Results

Table A16.1: Summary of badger monitoring surveys results – July/August 2020

Sett/Date	22.07.2020	27.07.2020	29.07.2020	03.08.2020	05.08.2020	12.08.2020	17.08.2020	Classification
Sett 1 (1-4)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 2 (1-10)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 3 (1-6)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 4 (1-2)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 5 (1-2)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 6 (1-3)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 7 (1)	N/A	A	A	NA	NA	A	A	A (outlier)
Sett 7 (2-4)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 8 (1-7, 9-14, 16-17, 19-26, 28-28, 40)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 8 (8, 18, 27, 39, 41)	N/A	NA	A (S)	NA	NA	NA	NA	A (main)
Sett 8 (8, 15, 39)	N/A	NA	NA	NA	NA	A (S)	NA	A (main)
Sett 8 (8, 9, 44)	N/A	NA	NA	NA	NA	NA	A (S)	A (main)
Sett 9 (1-3)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 10 (1, 3, 5)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 10 (2)	N/A	NA	A	NA	NA	NA	NA	A (subsidiary)
Sett 10 (4)	N/A	A	NA	A(S)	A	A(S)	NA	A (subsidiary)
Sett 11 (1-3)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 12 (1-2)	N/A	NA	NA	NA	NA	NA	NA	NA
Sett 13 (1)	Identified post monitoring period – Considered Active							

Key: A = Active; NA=No Activity; F=Footprint; S=Sticks Pushed Down; R=Rabbit; V = Fox; and B=Bedding

Blue = Active; White = Inactive

Table A16.2: Summary of badger scoping surveys results – April 2021

Sett number	Number of entrances	Sett type	Notes
1	4	Outlier	All entrances filled with leaf litter (disused)
2	10	Old disused subsidiary	All entrances filled with leaf litter (disused)
3	6	Old disused subsidiary	All entrances filled with leaf litter (disused)
4	2	Outlier	All entrances filled with leaf litter (disused)
5	2	Outlier	All entrances filled with leaf litter (disused)
6	3	Outlier	All entrances filled with leaf litter (disused)
7	4	Outlier	Entrances 2, 3 & 4 filled with leaf litter (partially used); entrance 1 had fresh spoil and badger fur in entrance (well used)
8	43	Main	Flags gone on most of the holes so no numbers. 33 holes were free from debris and had fresh spoil (well used). Of these, 11 showed signs of recent use such as fresh bedding and badger hairs. The other 10 entrances were filled with leaf litter (partially used)
9	3	Outlier	All entrances filled with leaf litter (disused)
10	5	Subsidiary	Entrances 1, 2, & 5 filled with leaf litter (partially used); entrances 2 & 4 had fresh spoil and badger fur was found in entrance 4 (well used)
11	3	Outlier	All entrances filled with leaf litter (disused)
12	2	Outlier	Both entrances filled with leaf litter (disused)
13	2	Outlier	Both entrances had fresh spoil (well used)

Appendix 15c – Badger Sett Location Plan

